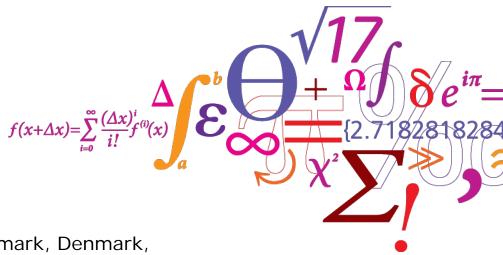


From Nanopore DNA Sequencing to

Organic Ices

for 3D electron beam patterning



Anpan Han, 韩安磐

Danchip/CEN, Technical University of Denmark, Denmark,

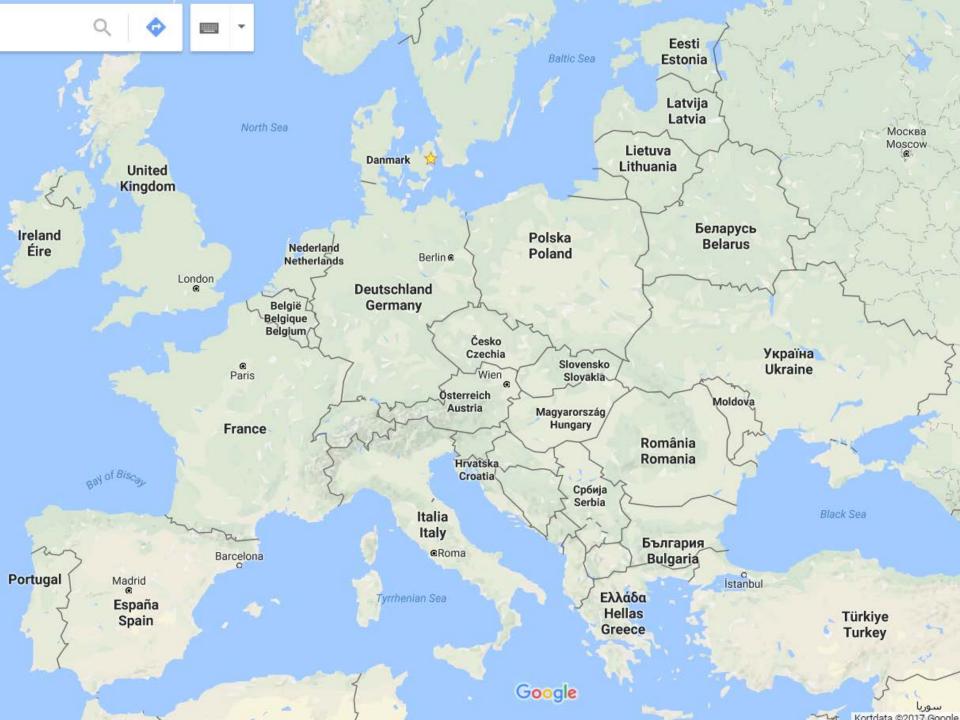
DTU Danchip National Center for Micro- and Nanofabrication **DTU Cen** Center for Electron Nanoscopy

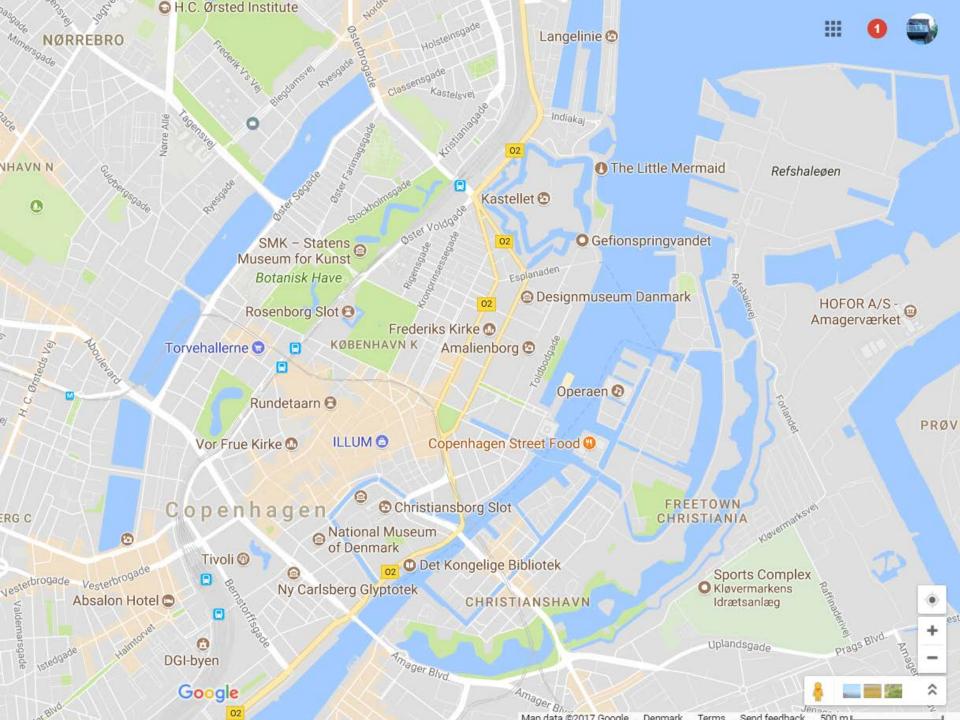
Content (75 + 10 min)

- Denmark, DTU, Danchip and CEN (5 min)
- Nanopore DNA sequencing (6 min)
- Ice lithography (15 min)
- Quiz (5 min), Break (10 min)

 $\overset{\wedge}{\searrow}$

- Organic ice resists for 3D nanolithography (15 min)
- Future research: 3D-OPE (5 min)
- Group work: 25 min: Please help me!



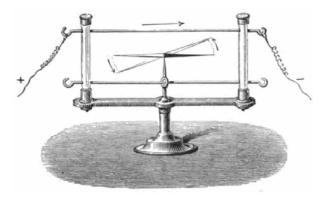




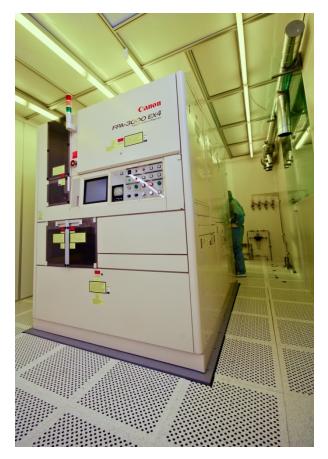
Technical University of Denmark

- Established 1829 by HC Oersted
- 1339 Faculty and Researcher
- 1540 Administrative and technical staff
- 6800 Undergraduate and Master student
- 1201 PhD students
- 26 Departments
- 30 min drive from Copenhagen Airport









150 nm half pitch 80 wafers/hour



300 kV aberration corrected environmental TEM



Cleanroom



Organic Ice

for 3D electron beam patterning ?!

How did you get this "seemingly implausible (SI)" idea?



HISTORICAL PERSPECTIVE

2016 nature biotechnology

\sim

Three decades of nanopore sequencing

David Deamer¹, Mark Akeson¹ & Daniel Branton²

A long-held goal in sequencing has been to use a voltage-biased nanoscale pore in a membrane to measure the passage of a linear, single-stranded (ss) DNA or RNA molecule through that pore. With the development of enzyme-based methods that ratchet polynucleotides through the nanopore, nucleobaseby-nucleobase, measurements of changes in the current through the pore can now be decoded into a DNA sequence using an algorithm. In this Historical Perspective, we describe the key steps in nanopore strand-sequencing, from its earliest conceptualization more than 25 years ago to its recent commercialization and application.

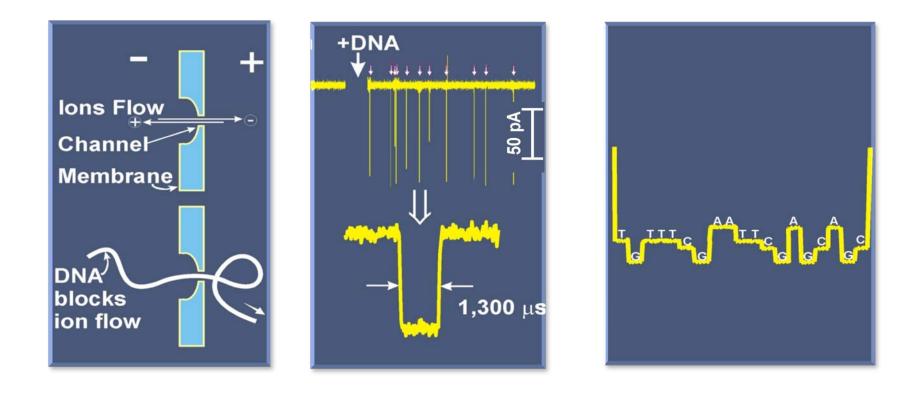
Nanopore sequencing (Box 1 and Fig. 1) has its origins in several laboratories during the 1980s (Fig. 2). In 1989, one of us (D.D.) jotted a seemingly implausible idea in his notebook (Fig. 3), suggesting that it might be possible to sequence a single strand of DNA being drawn through a membrane's nanoscopic pore by electrophoresis. Around the same time, George Church's interest in scaling up DNA sequencing for

and together with Sergey Bezrukov at the US National Institutes of Health (NIH), had established the conditions that were necessary to avoid spontaneous gating (pore closure) of the α -hemolysin channel³. This was important, because spontaneous gating would have hindered, or confused, observations of nucleic acid translocation through the nanopore. Kasianowicz was also collaborating with Bezrukov to investigate the effect of polyethylene glycol on pore conductance and, consistent with earlier reports², found that a pore radius of ~1.1 nm accounted for their results⁴.

In initial experiments, D.D. and Kasianowicz worked with a single α -hemolysin channel inserted into a lipid bilayer that separated two buffered KCl-filled compartments. RNA homopolymers (polyuridylic or polyadenylic acid) were then added to the *cis* side of the membrane. Immediately after addition of the polymers, the first encouraging results were observed. When a positive voltage bias greater than 80 mV was applied to the *trans* compartment, numerous blockades—transient, millisecond time-scale reductions of the ionic current through the α -hemolysin channel—appeared. No blockades were detected when the *trans* compartment was negatively biased. This was expected because the polyanionic RNA in the *cis* chamber would be inhibited from entering the nanopore when the *trans* side was negative. These preliminary results were consistent with the hypothesis that a voltage applied across the



Seemingly implausible Nanopore DNA sequencing idea (1989)





2018 Oxford Nanopore Technologies, 4th Gen DNA sequencing



nature biotechnology

OPEN

Linear assembly of a human centromere on the Y chromosome

Miten Jain^{1,5}, Hugh E Olsen^{1,5}, Daniel J Turner², David Stoddart², Kira V Bulazel³, Benedict Paten¹, David Haussler¹, Huntington F Willard^{3,4}, Mark Akeson¹ & Karen H Miga^{1,3}

The human genome reference sequence remains incomplete owing to the challenge of assembling long tracts of nearidentical tandem repeats in centromeres. We implemented a nanopore sequencing strategy to generate high-quality reads that span hundreds of kilobases of highly repetitive DNA in a human Y chromosome centromere. Combining these data with short-read variant validation, we assembled and characterized the centromeric region of a human Y chromosome.

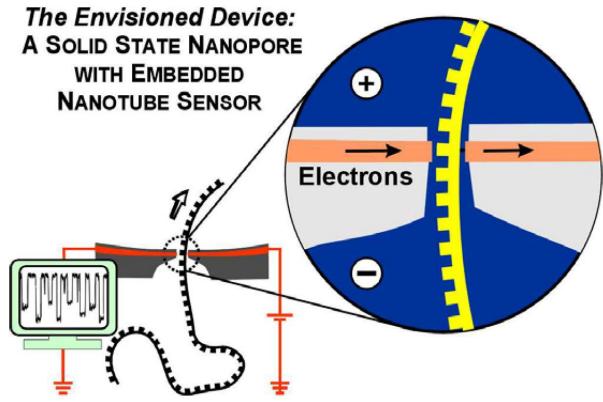
Centromeres facilitate spindle attachment and ensure proper chromosome segregation during cell division. Normal human centromeres are enriched with AT-rich ~171-bp tandem repeats known as alpha satellite DNA¹ Most alpha satellite DNAs are organized into higher longboard strategy, we linearize the circular BAC with a single cut site, then add sequencing adaptors (Fig. 1a). The BAC DNA passes through the pore, resulting in complete, end-to-end sequence coverage of the entire insert. Plots of read length versus megabase yield revealed an increase in megabase yield for full-length BAC DNA sequences (Fig. 1b and Supplementary Fig. 2). We present more than 3,500 full-length '1D' reads (that is, one strand of the DNA is sequenced) from ten BACs (two control BACs from Xq24 and Yp11.2; eight BACs in the DYZ3 locus⁹; Supplementary Table 1).

Correct assembly across the centromeric locus requires overlap among a few sequence variants, meaning that accuracy of base-calls is important. Individual reads (MinION R9.4 chemistry, Albacore v1.1.1) provide insufficient sequence identity (median alignment identity of 84.8% for control BAC, RP11-482A22 reads) to ensure correct repeat assembly¹⁰. To improve overall base quality, we produced a consensus sequence from 10 iterations of 60 randomly sampled alignments of full-length 1D reads that spanned the full insert length for each BAC (**Fig. 1c**). To polish sequences, we realigned full-length nanopore reads to each BAC-derived consensus (99.2% observed for control BAC, RP11-482A22; and an observed range of 99.4–99.8% for vector sequences in DYZ3-containing BACs). To provide a truth set of array sequence variants and to evaluate any inherent nanopore sequence biases, we used Illumina BAC resequencing (Online Methods). We used eight BAC-polished sequences (e.g., 209 kb for





Envisioned (SI) Nanopore DNA sequencing device 2005



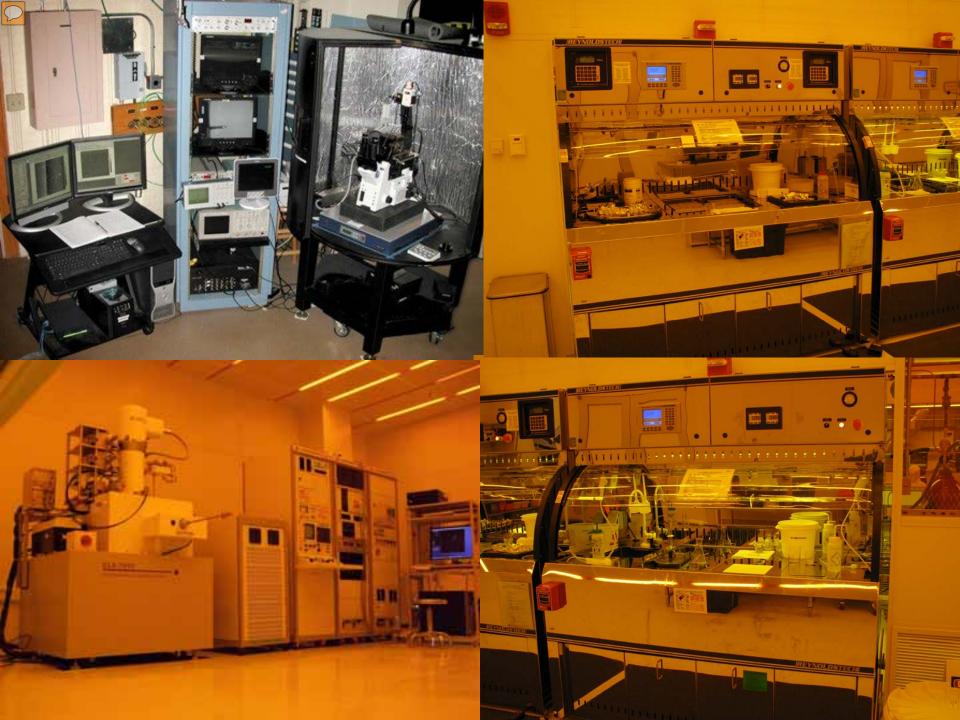


Dan Branton Mole. Bio. Harvard



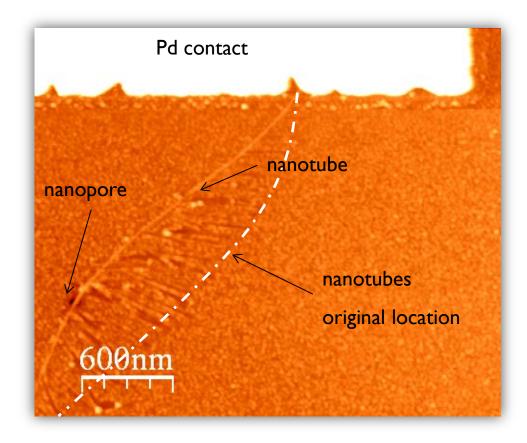
Jene Golovchenko SEAS, Physics, Harvard

Electrical contacts to carbon nanotubes by electron beam lithography





Aligning nanotube with nanopore 2006-2007







I have this great (SI) idea!

Lets use water ice as an e-beam resist!

Water is so much cleaner! I used to study proton - water ice interactions at Bell Labs.

I joined the group in 2008.



Dan Branton



Jene Golovchenko



Ice Lithography for Nanodevices



Anpan Han, Dimitar Vlassarev, Jenny Wang, Jene A. Golovchenko, Daniel Branton

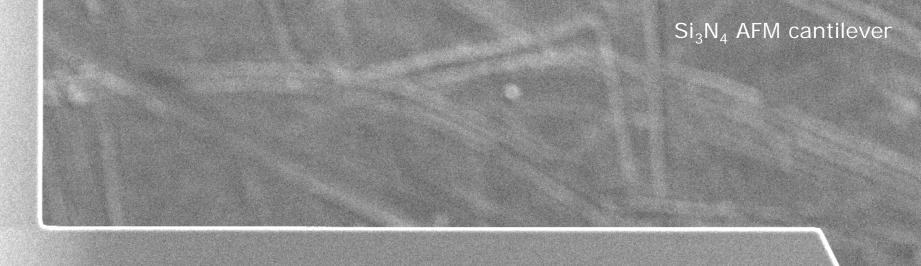
DTU Danchip CEN, Technical University of Denmark

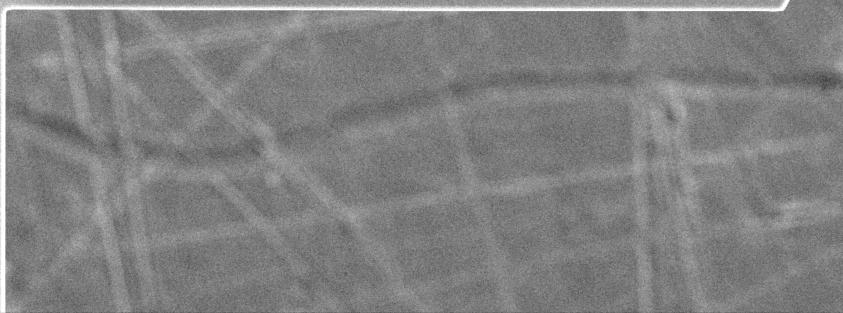
16 April 2018



We can pattern on non-planar samples!

Han et al. 2012, Nano Letters

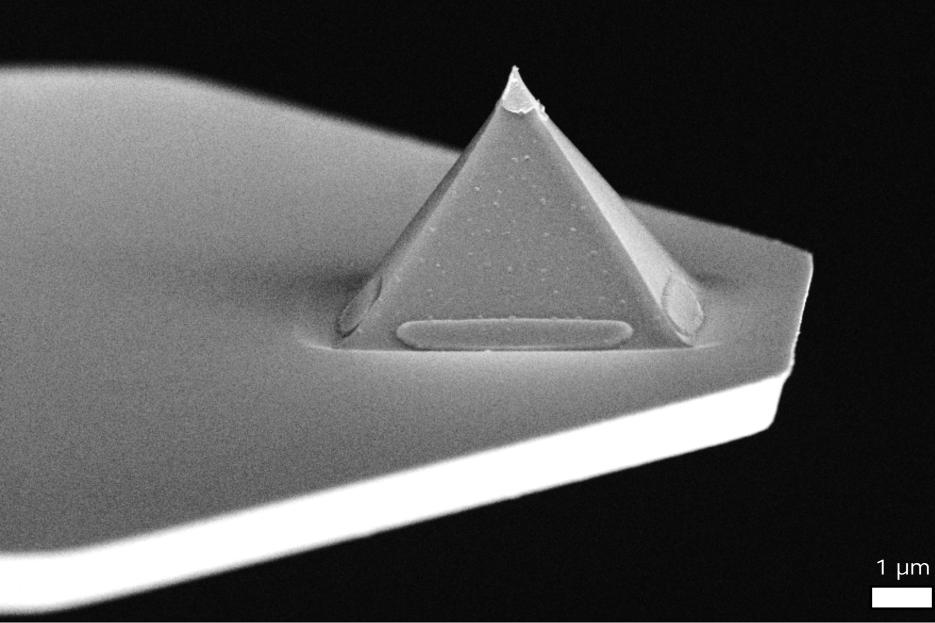




10µm JEOL

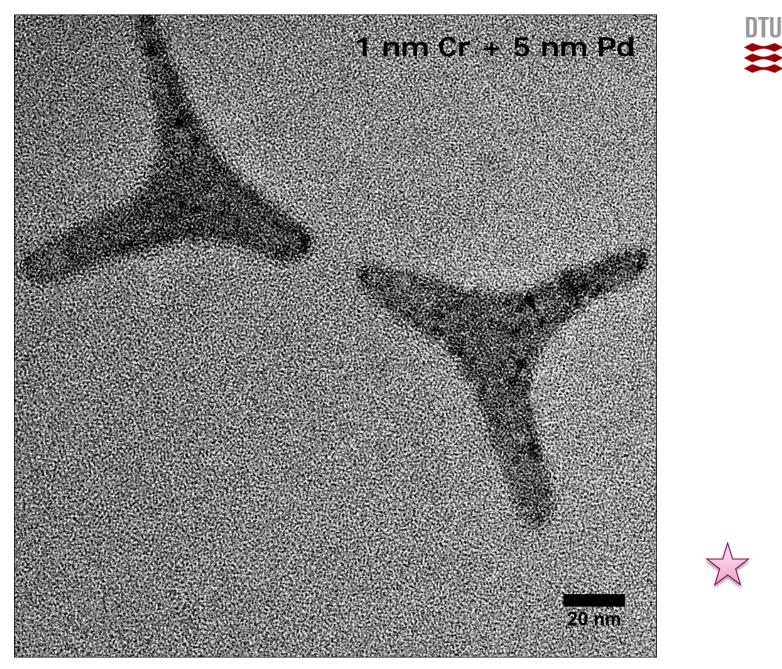


Metal coated apex: 1 nm Ti/ 20 nm Au

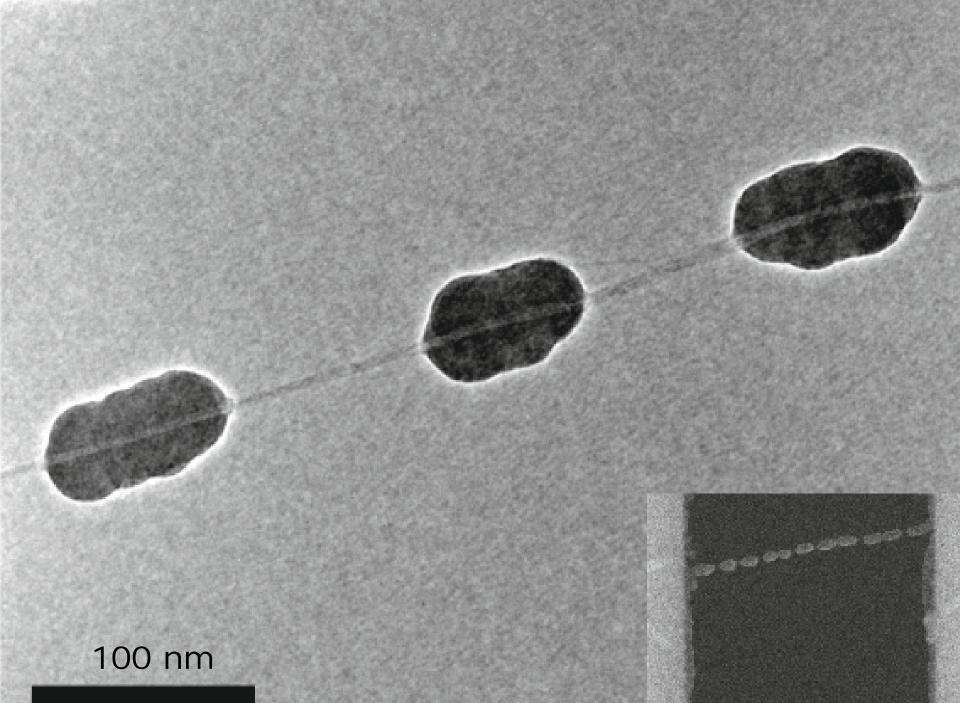


TEM grid, Si₃N₄ membranes, 20-60 nm thin



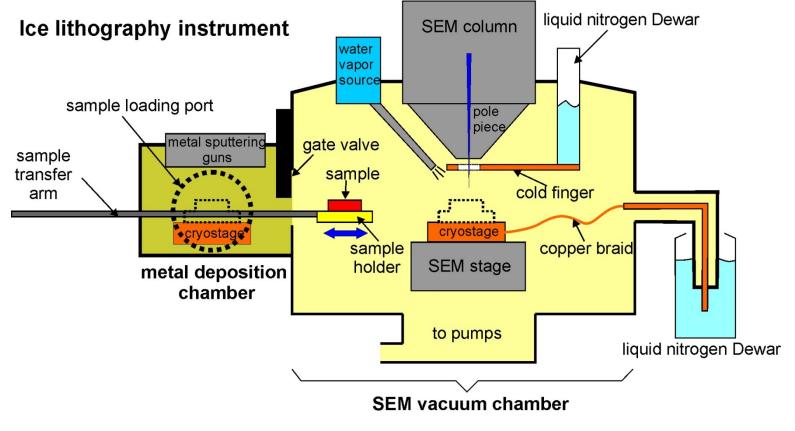






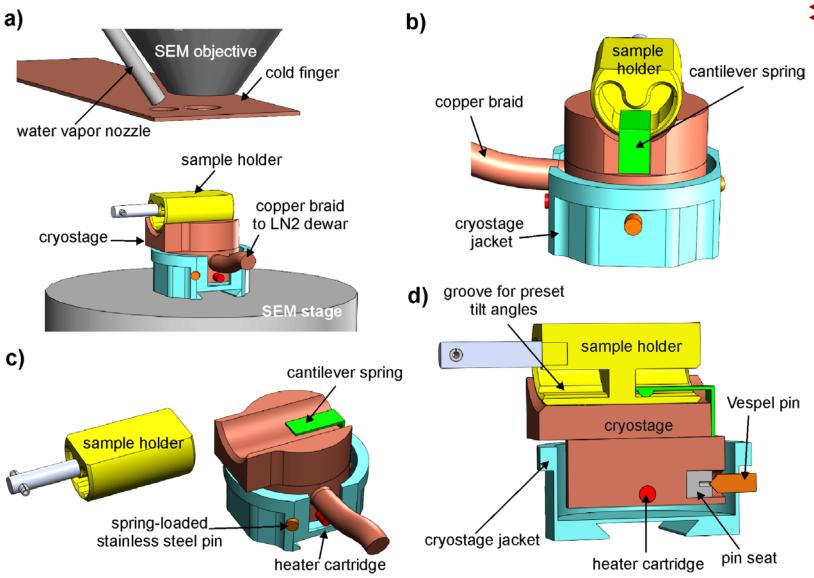


ice lithography instrument

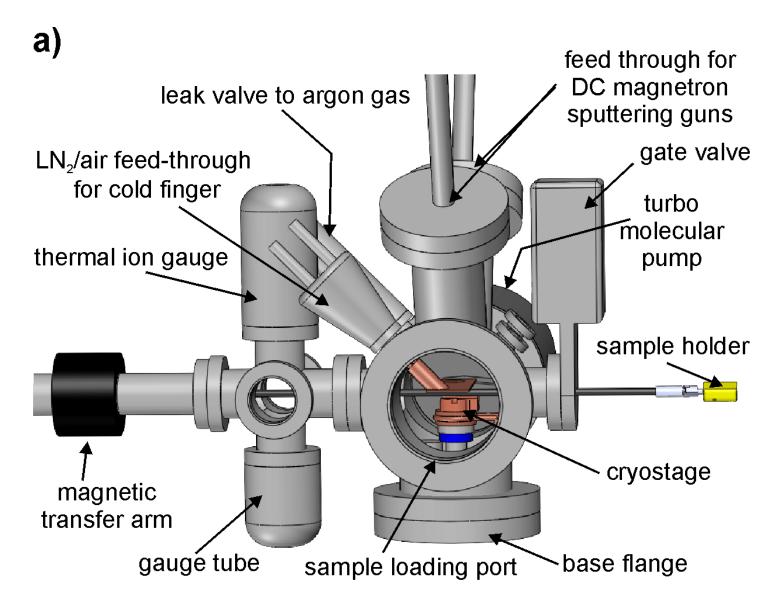


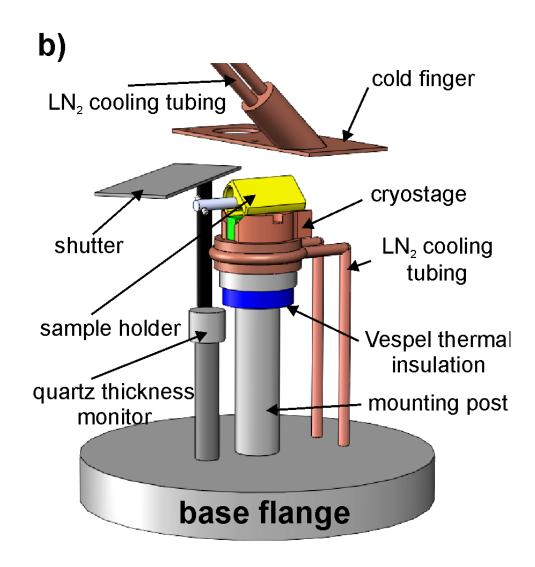
Han et al. 2011 Rev. Sci. Inst

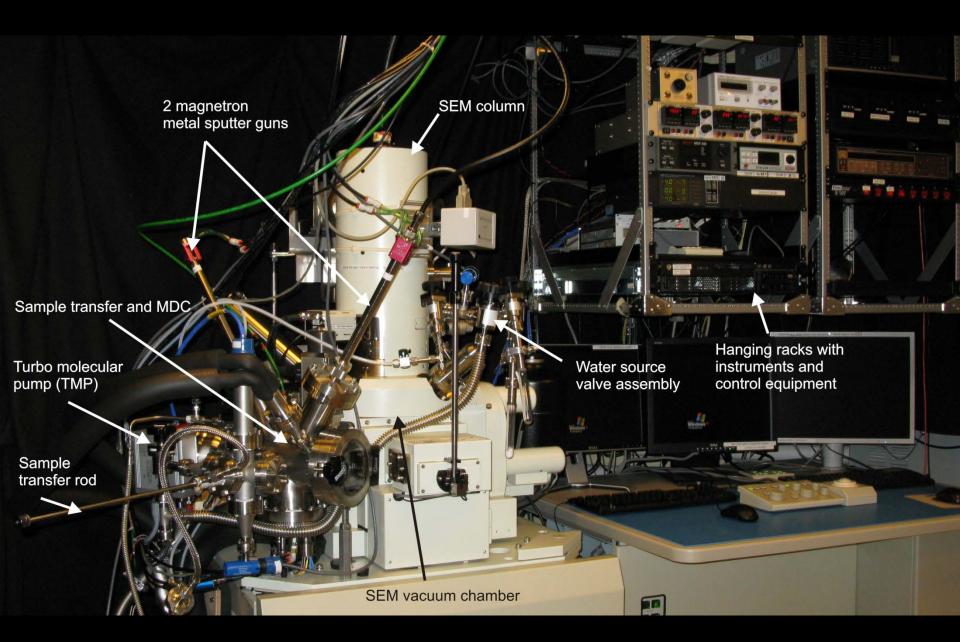














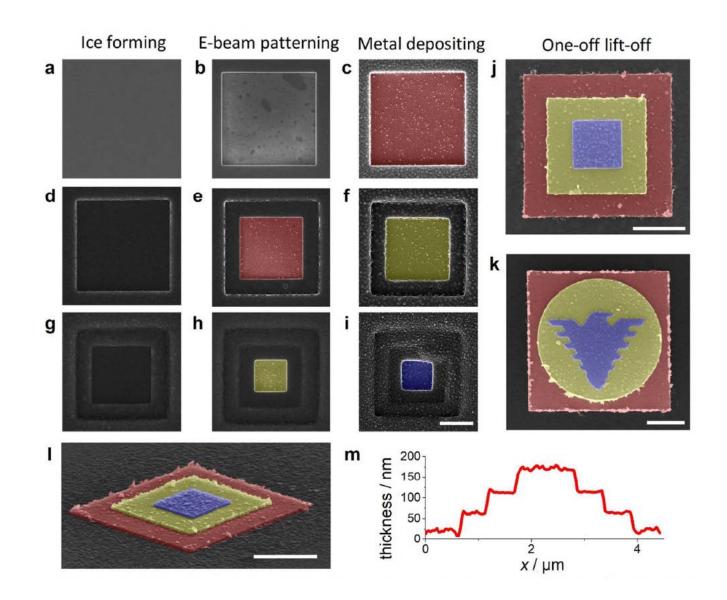


Subscriber access provided by DTU Library

Communication

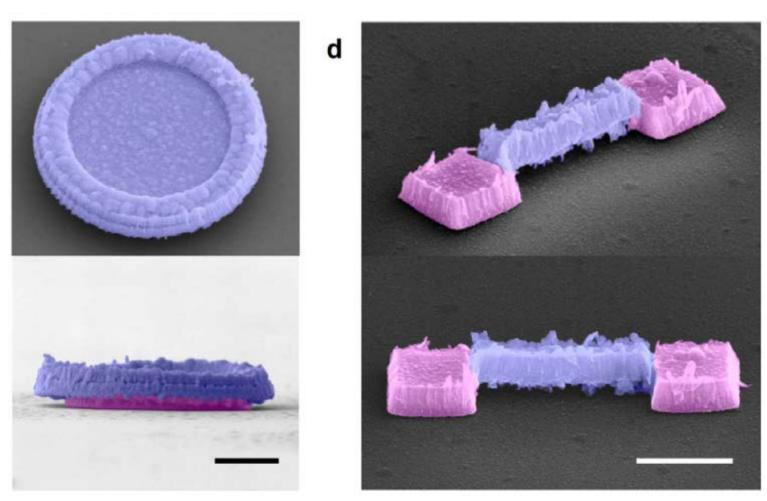
Three-dimensional in situ electron beam lithography using water ice

Yu Hong, Ding Zhao, Dongli Liu, Binze Ma, Guangnan Yao, Qiang Li, Anpan Han, and Min Qiu Nano Lett., Just Accepted Manuscript • DOI: 10.1021/acs.nanolett.8b01857 • Publication Date (Web): 25 Jun 2018 Downloaded from http://pubs.acs.org on July 2, 2018













Advantages

- Non-planar and extremely fragile substrates
- High resolution: sub-10 nm
- Rapid and resource efficient: prototype in 3 h
- Clean: no resist contamination
- Fabrication of nanodevices



Draw backs

- All processes at cryogenic temperatures
- High critical dose: 1C/cm2
- Advanced instrument, high cost

Better ices?



Quiz

- 1. What does SI in the presentation stand for?
- 2. How many authors on the "3 decades of Nanopore DNA sequencing" paper?
- 3. How is "scum" removed?
- 4. How many nm are the smallest patterns made by ice lithography?
- 5. What is the critical dose for water ice resist?
- 6. Advantages of ice lithography? The more the better.

Answers

- 1. What does SI in the presentation stand for? Seemingly implausible
- How many authors on the "3 decades of Nanopore DNA sequencing" paper?
 3
- 3. How is "scum" removed?
- 4. How many nm are the smallest patterns made by ice lithography? 6 nm
- 5. What is the critical dose for water ice resist? 1 C/cm2
- 6. Advantages of ice lithography? The more the better.

Oxygen plasma



10 min break, and bring your laptops



Seemingly implausible nanoscale 3D print of functional materials?

OVERVIEW OF THE 5 BRANCHES OF CHEMISTRY

- 1. Organic Chemistry The study of carbon and its compounds; the study of the chemistry of life.
- 2. **Inorganic Chemistry** The study of compounds not-covered by organic chemistry; the study of inorganic compounds or compounds which do not contain a C-H bond. Many inorganic compounds are those which contain metals.
- 3. **Analytical Chemistry** The study of the chemistry of matter and the development of tools used to measure properties of matter.
- 4. **Physical Chemistry** The branch of chemistry that applies physics to the study of chemistry. Commonly this includes the applications of thermodynamics and quantum mechanics to chemistry.
- 5. **Biochemistry** This is the study of chemical processes that occur inside of living organisms.



Branches of Organic Chemistry

- Metalorganic chemistry
- Polymer chemistry (dielectrics)
- Semiconducting organic chemistry
- and many more.



Acknowledgements at Harvard Uni.



Dan Branton, Jene Golovchenko, Lene Hau. Dimitar Vlassarev, Jenny Wang, Aaron Kuan.



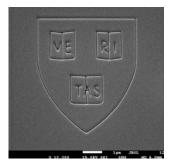
John Chervinski, Peter Frisella, Ray Aubut.

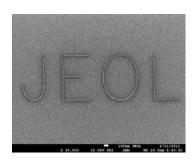


H National Human Genome Research Institute

CARL§BERGFONDET CARL§BERG FOUNDATION

















JEOL USA.



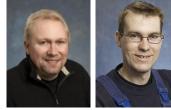
Acknowledgements for DTU ice activities

- Project: William Tiddi, Anna Elsukova, Liu Pei, Hoa Thanh Le, Marco Belegia
- Technical staff at Danchip CEN: Roy Cork, Jonas M. Lindhart, Anders Gregersen, Jakob B Wagner.
- Technical staff at DTU Risø: Michael Nimb; Troels Fel d, Johnny Egtved Jørgensen
- Management: Jörg Hübner, Flemming Jensen, Anders M. Jørgensen
- Zeiss R&D: Camille Stebler
 - THE VELUX FOUNDATIONS





















Group work: 3-4 people in each group (25 min)

- Killer application? What would you 3D print? (15 min)
- Present your killer application to the class.
 - 2 min for each group,
 - 2-3 slides
- Vote for best killer application measured in impact:
 - New needs, saving lives, market size, live better and longer, solve big problems
- Winners shares a gift from Denmark.